

# Package ‘doBy’

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**Title** Groupwise Statistics, LSmeans, Linear Estimates, Utilities

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**Description** Utility package containing:

- 1) Facilities for working with grouped data: 'do' something to data stratified 'by' some variables.
- 2) LSmeans (least-squares means), general linear estimates.
- 3) Restrict functions to a smaller domain.
- 4) Miscellaneous other utilities.

**Encoding** UTF-8

**VignetteBuilder** knitr

**ZipData** no

**LazyData** true

**LazyDataCompression** xz

**License** GPL (>= 2)

**Depends** R (>= 3.6.0), methods

**Imports** broom, Deriv, dplyr, ggplot2, MASS, Matrix, magrittr,  
microbenchmark, pbkrtest (>= 0.4-8.1), tibble

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<code>.rhsf2list</code>	<i>Convert right hand sided formula to a list</i>
-------------------------	---

---

**Description**

Convert right hand sided formula to a list

**Usage**

`.rhsf2list(f)`

**Arguments**

f                   A right hand sided formula

---

beets	<i>beets data</i>
-------	-------------------

---

**Description**

Yield and sugar percentage in sugar beets from a split plot experiment. Data is obtained from a split plot experiment. There are 3 blocks and in each of these the harvest time defines the "whole plot" and the sowing time defines the "split plot". Each plot was 25m<sup>2</sup> and the yield is recorded in kg. See 'details' for the experimental layout.

**Usage**

beets

**Format**

The format is: chr "beets"

**Details**

Experimental plan

Sowing times	1	4. april
	2	12. april
	3	21. april
	4	29. april
	5	18. may
Harvest times	1	2. october
	2	21. october

Plot allocation:

	Block 1	Block 2	Block 3	
	+-----+ -----+ -----+			
Plot 1-15	1 1 1 1 1	2 2 2 2 2	1 1 1 1 1	Harvest time
	3 4 5 2 1	3 2 4 5 1	5 2 3 4 1	Sowing time
	-----+ -----+ -----+			
Plot 16-30	2 2 2 2 2	1 1 1 1 1	2 2 2 2 2	Harvest time
	2 1 5 4 3	4 1 3 2 5	1 4 3 2 5	Sowing time
	+-----+ -----+ -----+			

**References**

Ulrich Halekoh, Søren Højsgaard (2014)., A Kenward-Roger Approximation and Parametric Bootstrap Methods for Tests in Linear Mixed Models - The R Package pbkrtest., Journal of Statistical Software, 58(10), 1-30., <https://www.jstatsoft.org/v59/i09/>

**Examples**

```
data(beets)

beets$bh <- with(beets, interaction(block, harvest))
summary(aov(yield ~ block + sow + harvest + Error(bh), beets))
summary(aov(sugpct ~ block + sow + harvest + Error(bh), beets))
```

---

breastcancer

*Gene expression signatures for p53 mutation status in 250 breast cancer samples*

---

**Description**

Perturbations of the p53 pathway are associated with more aggressive and therapeutically refractory tumours. We preprocessed the data using Robust Multichip Analysis (RMA). Dataset has been truncated to the 1000 most informative genes (as selected by Wilcoxon test statistics) to simplify computation. The genes have been standardised to have zero mean and unit variance (i.e. z-scored).

**Usage**

breastcancer

**Format**

A data frame with 250 observations on 1001 variables. The first 1000 columns are numerical variables; the last column (named code) is a factor with levels case and control.

**Details**

The factor code defines whether there was a mutation in the p53 sequence (code=case) or not (code=control).

**Source**

Dr. Chris Holmes, c.holmes at stats dot. ox . ac .uk

**References**

Miller et al (2005, PubMed ID:16141321)

**Examples**

```
data(breastcancer)
bc <- breastcancer
pairs(bc[,1:5], col=bc$code)

train <- sample(1:nrow(bc), 50)
table(bc$code[train])
## Not run:
library(MASS)
z <- lda(code ~ ., data=bc, prior = c(1,1)/2, subset = train)
pc <- predict(z, bc[-train, ])$class
pc
bc[-train, "code"]
table(pc, bc[-train, "code"])

## End(Not run)
```

---

budworm

*budworm data*

---

**Description**

Effect of Insecticide on survival of tobacco budworms number of killed budworms exposed to an insecticidepp mortality of the moth tobacco budworm 'Heliothis virescens' for 6 doses of the pyrethroid trans-cypermethrin differentiated with respect to sex

**Usage**

budworm

**Format**

This data frame contains 12 rows and 4 columns:

**sex:** sex of the budworm

**dose:** dose of the insecticide trans-cypermethrin in [ $\mu\text{g}$ ]

**ndead:** budworms killed in a trial

**ntotal:** total number of budworms exposed per trial

**Source**

Collet, D. (1991) Modelling Binary Data, Chapman & Hall, London, Example 3.7

**References**

Venables, W.N; Ripley, B.D.(1999) Modern Applied Statistics with S-Plus, Heidelberg, Springer, 3rd edition, chapter 7.2

**Examples**

```
data(budworm)

## function to calculate the empirical logits
empirical.logit<- function(y, n) {
  el <- log((y + 0.5) / (n - y + 0.5))
  el
}

# plot the empirical logits against log-dose

log.dose <- log(budworm$dose)
emp.logit <- empirical.logit(budworm$ndead, budworm$ntotal)
plot(log.dose, emp.logit, type='n', xlab='log-dose', ylab='empirical logit')
title('budworm: empirical logits of probability to die ')
male <- budworm$sex=='male'
female <- budworm$sex=='female'
lines(log.dose[male], emp.logit[male], type='b', lty=1, col=1)
lines(log.dose[female], emp.logit[female], type='b', lty=2, col=2)
legend(0.5, 2, legend=c('male', 'female'), lty=c(1, 2), col=c(1, 2))

## Not run:
* SAS example;
data budworm;
infile 'budworm.txt' firstobs=2;
input sex dose ndead ntotal;
run;

## End(Not run)
```

---

`by-lapply`*Formula based version of lapply and sapply*

---

**Description**

This function is a wrapper for calling `lapply` on the list resulting from first calling `splitBy`.

**Usage**

```
lapply_by(data, formula, FUN, ...)
```

```
lapplyBy(formula, data = parent.frame(), FUN, ...)
```

```
sapply_by(data, formula, FUN, ..., simplify = TRUE, USE.NAMES = TRUE)
```

```
sapplyBy(  
  formula,  
  data = parent.frame(),  
  FUN,  
  ...,  
  simplify = TRUE,  
  USE.NAMES = TRUE  
)
```

**Arguments**

<code>data</code>	A dataframe.
<code>formula</code>	A formula describing how data should be split.
<code>FUN</code>	A function to be applied to each element in the splitted list, see 'Examples' below.
<code>...</code>	optional arguments to <code>FUN</code> .
<code>simplify</code>	Same as for 'sapply'
<code>USE.NAMES</code>	Same as for 'sapply'

**Value**

A list.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[splitBy](#), [split\\_by](#)

**Examples**

```

fun <- function(x) range(x$uptake)
lapplyBy(~Treatment + Type, data=CO2, FUN=fun)
sapplyBy(~Treatment + Type, data=CO2, FUN=fun)

# Same as
lapply(splitBy(~Treatment + Type, data=CO2), FUN=fun)

```

---

by-lmby

---

*List of lm objects with a common model*


---

**Description**

The data is split into strata according to the levels of the grouping factors and individual lm fits are obtained for each stratum.

**Usage**

```

lm_by(data, formula, id = NULL, ...)

lmBy(formula, data, id = NULL, ...)

```

**Arguments**

data	A dataframe
formula	A linear model formula object of the form $y \sim x_1 + \dots + x_n \mid g_1 + \dots + g_m$ . In the formula object, $y$ represents the response, $x_1, \dots, x_n$ the covariates, and the grouping factors specifying the partitioning of the data according to which different lm fits should be performed.
id	A formula describing variables from data which are to be available also in the output.
...	Additional arguments passed on to <code>lm()</code> .

**Value**

A list of lm fits.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>



## Examples

```
bb <- lmBy(1 / uptake ~ log(conc) | Treatment, data=C02)
coef(bb)

fitted(bb)
residuals(bb)

summary(bb)
coef(summary(bb))
coef(summary(bb), simplify=TRUE)
```

---

by-order

*Ordering (sorting) rows of a data frame*

---

## Description

Ordering (sorting) rows of a data frame by the certain variables in the data frame. This function is essentially a wrapper for the `order()` function - the important difference being that variables to order by can be given by a model formula.

## Usage

```
order_by(data, formula)

orderBy(formula, data)
```

## Arguments

data	A dataframe
formula	The right hand side of a formula

## Details

The sign of the terms in the formula determines whether sorting should be ascending or decreasing; see examples below

## Value

The ordered data frame

## Author(s)

Søren Højsgaard, <sorenh@math.aau.dk> and Kevin Wright

## See Also

[transformBy](#), [transform\\_by](#), [splitBy](#), [split\\_by](#)

## Examples

```
orderBy(~ conc + Treatment, C02)
## Sort decreasingly by conc
orderBy(~ - conc + Treatment, C02)
```

---

by-sample

*Sampling from a data frame*

---

## Description

A data frame is split according to some variables in a formula, and a sample of a certain fraction of each is drawn.

## Usage

```
sample_by(data, formula, frac = 0.1, replace = FALSE, systematic = FALSE)
```

```
sampleBy(
  formula,
  frac = 0.1,
  replace = FALSE,
  data = parent.frame(),
  systematic = FALSE
)
```

## Arguments

data	A data frame.
formula	A formula defining the grouping of the data frame.
frac	The part of data to be sampled.
replace	Is the sampling with replacement.
systematic	Should sampling be systematic.

## Details

If systematic=FALSE (default) then frac gives the fraction of data sampled. If systematic=TRUE and frac=.2 then every 1/.2 i.e. every 5th observation is taken out.

## Value

A dataframe.

## See Also

[orderBy](#), [order\\_by](#), [splitBy](#), [split\\_by](#), [summaryBy](#), [summary\\_by](#), [transformBy](#), [transform\\_by](#)

**Examples**

```
data(dietox)
sampleBy(formula = ~ Evit + Cu, frac=.1, data = dietox)
```

by-split

*Split a data frame***Description**

Split a dataframe according to the levels of variables in the dataframe. The variables to split by can be given as a formula or as a character vector.

**Usage**

```
split_by(data, formula, drop = TRUE)

splitBy(formula, data = parent.frame(), drop = TRUE)

## S3 method for class 'splitByData'
head(x, n = 6L, ...)

## S3 method for class 'splitByData'
tail(x, n = 6L, ...)
```

**Arguments**

data	A data frame
formula	Variables to split data frame by, as ‘as.quoted’ variables, a formula or character vector.
drop	Logical indicating if levels that do not occur should be dropped. Deprecated; levels that do not occur are ignored.
x	An object.
n	A single integer. If positive or zero, size for the resulting object: number of elements for a vector (including lists), rows for a matrix or data frame or lines for a function. If negative, all but the "n" last/first number of elements of "x".
...	Arguments to be passed to or from other methods.

**Value**

A list of dataframes.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[orderBy](#), [order\\_by](#), [summaryBy](#), [summary\\_by](#), [transformBy](#), [transform\\_by](#)

**Examples**

```
data(dietox, package="doBy")
splitBy(formula = ~Evit + Cu, data = dietox)
splitBy(formula = c("Evit", "Cu"), data = dietox)

splitBy(~Treatment + Type, data=C02)
splitBy(c("Treatment", "Type"), data=C02)

x <- splitBy(~Treatment, data=C02)
head(x)
tail(x)
```

---

by-subset

*Finds subsets of a dataframe which is split by variables in a formula.*

---

**Description**

A data frame is split by a formula into groups. Then subsets are found within each group, and the result is collected into a data frame.

**Usage**

```
subset_by(data, formula, subset, select, drop = FALSE, join = TRUE, ...)
```

```
subsetBy(
  formula,
  subset,
  data = parent.frame(),
  select,
  drop = FALSE,
  join = TRUE,
  ...
)
```

**Arguments**

data	A data frame.
formula	A right hand sided formula or a character vector of variables to split by.
subset	logical expression indicating elements or rows to keep: missing values are taken as false.
select	expression, indicating columns to select from a data frame.
drop	passed on to [ indexing operator.

`join` If FALSE the result is a list of data frames (as defined by 'formula'); if TRUE one data frame is returned.

`...` further arguments to be passed to or from other methods.

**Value**

A data frame.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[splitBy](#), [split\\_by](#)

**Examples**

```
data(dietox)
subsetBy(~Evit, Weight < mean(Weight), data=dietox)
```

---

by-summary

*Function to calculate groupwise summary statistics*

---

**Description**

Function to calculate groupwise summary statistics, much like the summary procedure of SAS

**Usage**

```
summary_by(
  data,
  formula,
  id = NULL,
  FUN = mean,
  keep.names = FALSE,
  p2d = FALSE,
  order = TRUE,
  full.dimension = FALSE,
  var.names = NULL,
  fun.names = NULL,
  ...
)

summaryBy(
  formula,
  data = parent.frame(),
```

```

    id = NULL,
    FUN = mean,
    keep.names = FALSE,
    p2d = FALSE,
    order = TRUE,
    full.dimension = FALSE,
    var.names = NULL,
    fun.names = NULL,
    ...
  )

```

### Arguments

<code>data</code>	A data frame.
<code>formula</code>	A formula object, see examples below.
<code>id</code>	A formula specifying variables which data are not grouped by but which should appear in the output. See examples below.
<code>FUN</code>	A list of functions to be applied, see examples below.
<code>keep.names</code>	If TRUE and if there is only ONE function in FUN, then the variables in the output will have the same name as the variables in the input, see 'examples'.
<code>p2d</code>	Should parentheses in output variable names be replaced by dots?
<code>order</code>	Should the resulting dataframe be ordered according to the variables on the right hand side of the formula? (using <a href="#">orderBy</a> )
<code>full.dimension</code>	If TRUE then rows of summary statistics are repeated such that the result will have the same number of rows as the input dataset.
<code>var.names</code>	Option for user to specify the names of the variables on the left hand side.
<code>fun.names</code>	Option for user to specify function names to apply to the variables on the left hand side.
<code>...</code>	Additional arguments to FUN. This could for example be NA actions.

### Details

Extra arguments ('...') are passed onto the functions in FUN. Hence care must be taken that all functions in FUN accept these arguments - OR one can explicitly write a functions which get around this. This can particularly be an issue in connection with handling NAs. See examples below. Some code for this function has been suggested by Jim Robison-Cox. Thanks.

### Value

A dataframe.

### Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[ave](#), [descStat](#), [orderBy](#), [order\\_by](#), [splitBy](#), [split\\_by](#), [transformBy](#), [transform\\_by](#)

**Examples**

```

data(dietox)
dietox12 <- subset(dietox,Time==12)

fun <- function(x){
  c(m=mean(x), v=var(x), n=length(x))
}

summaryBy(cbind(Weight, Feed) ~ Evit + Cu, data=dietox12,
          FUN=fun)

summaryBy(list(c("Weight", "Feed"), c("Evit", "Cu")), data=dietox12,
          FUN=fun)

## Computations on several variables is done using cbind( )
summaryBy(cbind(Weight, Feed) ~ Evit + Cu, data=subset(dietox, Time > 1),
          FUN=fun)

## Calculations on transformed data is possible using cbind( ), but
# the transformed variables must be named

summaryBy(cbind(lw=log(Weight), Feed) ~ Evit + Cu, data=dietox12, FUN=mean)

## There are missing values in the 'airquality' data, so we remove these
## before calculating mean and variance with 'na.rm=TRUE'. However the
## length function does not accept any such argument. Hence we get
## around this by defining our own summary function in which length is
## not supplied with this argument while mean and var are:

sumfun <- function(x, ...){
  c(m=mean(x, na.rm=TRUE, ...), v=var(x, na.rm=TRUE, ...), l=length(x))
}
summaryBy(cbind(Ozone, Solar.R) ~ Month, data=airquality, FUN=sumfun )

## Using '.' on the right hand side of a formula means to stratify by
## all variables not used elsewhere:

data(warpbreaks)
summaryBy(breaks ~ wool + tension, warpbreaks, FUN=mean)
summaryBy(breaks ~ ., warpbreaks, FUN=mean)
summaryBy(. ~ wool + tension, warpbreaks, FUN=mean)

```

**Description**

Function to make groupwise transformations of data by applying the transform function to subsets of data.

**Usage**

```
transform_by(data, formula, ...)
```

```
transformBy(formula, data, ...)
```

**Arguments**

data	A data frame
formula	A formula with only a right hand side, see examples below
...	Further arguments of the form tag=value

**Details**

The ... arguments are tagged vector expressions, which are evaluated in the data frame data. The tags are matched against names(data), and for those that match, the value replace the corresponding variable in data, and the others are appended to data.

**Value**

The modified value of the dataframe data.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[orderBy](#), [order\\_by](#), [summaryBy](#), [summary\\_by](#), [splitBy](#), [split\\_by](#)

**Examples**

```
data(dietox)
transformBy(~Pig, data=dietox, minW=min(Weight), maxW=max(Weight),
  gain=diff(range(Weight)))
```



---

by_scale	<i>Scale a dataframe or matrix</i>
----------	------------------------------------

---

**Description**

Split a dataframe into a list according to the levels of variables in the dataframe and scale the numeric variables in each dataframe in the list.

**Usage**

```
scaleBy(formula, data = parent.frame(), center = TRUE, scale = TRUE)
```

```
scale_by(data, formula, center = TRUE, scale = TRUE)
```

**Arguments**

formula	Variables to split data frame by, as ‘as.quoted’ variables, a formula or character vector.
data	A dataframe or matrix
center	Logical, should data be centered.
scale	Logical, should data be scaled.

**Value**

A list of objects of same class as ‘x’

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[orderBy](#), [order\\_by](#), [summaryBy](#), [summary\\_by](#), [transformBy](#), [transform\\_by](#)

**Examples**

```
scaleBy(~Species, data=iris, center=TRUE, scale=FALSE)
scaleBy(~1, data=iris, center=TRUE, scale=FALSE)

scale_by(iris, ~Species)
scale_by(iris, ~1)

## Not combine list of dataframes to one dataframe e.g. as:
a <- scale_by(iris, ~Species)
d <- do.call(rbind, a)
```

---

 carcass

*Lean meat contents of 344 pig carcasses*


---

**Description**

Measurement of lean meat percentage of 344 pig carcasses together with auxillary information collected at three Danish slaughter houses

**Usage**

carcass

**Format**

carcassall: A data frame with 344 observations on the following 17 variables.

weight Weight of carcass

lengthc Length of carcass from back toe to head (when the carcass hangs in the back legs)

lengthf Length of carcass from back toe to front leg (that is, to the shoulder)

lengthp Length of carcass from back toe to the pelvic bone

Fat02, Fat03, Fat11, Fat12, Fat13, Fat14, Fat16 Thickness of fat layer at different locations on the back of the carcass (FatXX refers to thickness at (or rather next to) rib no. XX. Notice that 02 is closest to the head

Meat11, Meat12, Meat13 Thickness of meat layer at different locations on the back of the carcass, see description above

LeanMeat Lean meat percentage determined by dissection

slhouse Slaughter house; a factor with levels slh1 and slh2.

sex Sex of the pig; a factor with levels castrate and female.

size Size of the carcass; a factor with levels normal and large. Here, 'normal' refers to carcass weight under 80 kg; 'large' refers to carcass weights between 80 and 110 kg.

**Details**

: Notice that there were slaughtered large pigs only at one slaughter house.

**Note**

carcass: Contains only the variables Fat11, Fat12, Fat13, Meat11, Meat12, Meat13, LeanMeat

**Source**

Busk, H., Olsen, E. V., Brøndum, J. (1999) Determination of lean meat in pig carcasses with the Autofom classification system, *Meat Science*, 52, 307-314

**Examples**

```
data(carcass)
head(carcass)
```

---

codstom	<i>Diet of Atlantic cod in the Gulf of St. Lawrence (Canada)</i>
---------	--

---

**Description**

Stomach content data for Atlantic cod (*Gadus morhua*) in the Gulf of St. Lawrence, Eastern Canada. Note: many prey items were of no interest for this analysis and were regrouped into the "Other" category.

**Usage**

```
codstom
```

**Format**

A data frame with 10000 observations on the following 10 variables.

`region` a factor with levels SGSL NGSL representing the southern and northern Gulf of St. Lawrence, respectively

`ship.type` a factor with levels 2 3 31 34 90 99

`ship.id` a factor with levels 11558 11712 136148 136885 136902 137325 151225 151935 99433

`trip` a factor with levels 10 11 12 179 1999 2 2001 20020808 3 4 5 6 7 8 88 9 95

`set` a numeric vector

`fish.id` a numeric vector

`fish.length` a numeric vector, length in mm

`prey.mass` a numeric vector, mass of item in stomach, in g

`prey.type` a factor with levels Ammodytes\_sp Argis\_dent Chion\_opil Detritus Empty Eualus\_fab Eualus\_mac Gadus\_mor Hyas\_aran Hyas\_coar Lebbeus\_gro Lebbeus\_pol Leptoctl\_mac Mallot\_vil Megan\_norv Ophiuroidea Other Paguridae Pandal\_bor Pandal\_mon Pasiph\_mult Sabin\_sept Sebastes\_sp Them\_abys Them\_comp Them\_lib

**Details**

Cod are collected either by contracted commercial fishing vessels (`ship.type` 90 or 99) or by research vessels. Commercial vessels are identified by a unique `ship.id`.

Either one research vessel or several commercial vessels conduct a survey (`trip`), during which a trawl, gillnets or hooked lines are set several times. Most trips are random stratified surveys (depth-based stratification).

Each trip takes place within one of the regions. The trip label is only guaranteed to be unique within a region and the `set` label is only guaranteed to be unique within a trip.

For each fish caught, the `fish.length` is recorded and the fish is allocated a `fish.id`, but the `fish.id` is only guaranteed to be unique within a set. A subset of the fish caught are selected for stomach analysis (stratified random selection according to fish length; unit of stratification is the set for research surveys, the combination `ship.id` and stratum for surveys conducted by commercial vessels, although strata are not shown in `codstom`).

The basic experimental unit in this data set is a cod stomach (one stomach per fish). Each stomach is uniquely identified by a combination of `region`, `ship.type`, `ship.id`, `trip`, `set`, and `fish.id`.

For each prey item found in a stomach, the species and mass of the prey item are recorded, so there can be multiple observations per stomach. There may also be several prey items with the same `prey.type` in the one stomach (for example many `prey.types` have been recoded `Other`, which produced many instances of `Other` in the same stomach).

If a stomach is empty, a single observation is recorded with `prey.type` `Empty` and a `prey.mass` of zero.

## Source

Small subset from a larger dataset (more stomachs, more variables, more `prey.types`) collected by D. Chabot and M. Hanson, Fisheries & Oceans Canada ([chabotd@dfo-mpo.gc.ca](mailto:chabotd@dfo-mpo.gc.ca)).

## Examples

```
data(codstom)
str(codstom)
# removes multiple occurrences of same prey.type in stomachs
codstom1 <- summaryBy(prey.mass ~
  region + ship.type + ship.id + trip + set + fish.id + prey.type,
  data = codstom,
  FUN = sum)

# keeps a single line per stomach with the total mass of stomach content
codstom2 <- summaryBy(prey.mass ~ region + ship.type + ship.id + trip + set + fish.id,
  data = codstom,
  FUN = sum)

# mean prey mass per stomach for each trip
codstom3 <- summaryBy(prey.mass.sum ~ region + ship.type + ship.id + trip,
  data = codstom2, FUN = mean)

## Not run:
# wide version, one line per stomach, one column per prey type
library(reshape)
codstom4 <- melt(codstom, id = c(1:7, 9))
codstom5 <- cast(codstom4,
  region + ship.type + ship.id + trip + set + fish.id + fish.length ~
  prey.type, sum)
k <- length(names(codstom5))
prey_col <- 8:k
out <- codstom5[,prey_col]
out[is.na(out)] <- 0
```

```
codstom5[,prey_col] <- out
codstom5$total.content <- rowSums(codstom5[, prey_col])

## End(Not run)
```

---

crimeRate

*crimeRate*

---

### Description

Crime rates per 100,000 inhabitants in states of the USA for different crime types in 1977.

### Usage

```
crimeRate
```

### Format

This data frame contains:

**State:** State of the USA

**Murder:** crime of murder

**Rape:**

**Robbery:**

**Assault:**

**Burglary:** residential theft

**Larceny:** unlawful taking of personal property (pocket picking)

**AutoTheft:**

### Examples

```
data(crimeRate)
```

---

`crime_rate`*crime\_rate*

---

**Description**

Crime rates per 100,000 inhabitants in states of the USA for different crime types in 1977.

**Usage**`crime_rate`**Format**

This data frame contains:

**Murder:** crime of murder

**Rape:**

**Robbery:**

**Assault:**

**Burglary:** residential theft

**Larceny:** unlawful taking of personal property (pocket picking)

**AutoTheft:**

**Examples**`data(crime_rate)`

---

`cropyield`*Yield from Danish agricultural production of grain and root crop.*

---

**Description**

Yield from Danish agricultural production of grain and root crop.

**Usage**`cropyield`

**Format**

A dataframe with 97 rows and 7 columns.

year From 1901 to 1997.

precip Milimeter precipitation.

yield Million feed units (se details).

area Area in 1000 ha for grains and root crop.

fertil 1000 tons fertilizer.

avgtmp1 Average temperature April-June (3 months).

avgtmp2 Average temperature July-October (4 months).

**Details**

A feed unit is the amount of energy in a kg of barley.

**References**

Danmarks statistik (Statistics Denmark).

---

data-mathmark

*Mathematics marks for students*

---

**Description**

The mathmark data frame has 88 rows and 5 columns.

**Usage**

```
data(mathmark)
```

**Format**

This data frame contains the following columns: mechanics, vectors, algebra, analysis, statistics.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**References**

David Edwards, An Introduction to Graphical Modelling, Second Edition, Springer Verlag, 2000

**Examples**

```
data(mathmark)
```

---

data-personality      *Personality traits*

---

**Description**

The 'peronality' dataframe has 240 rows and 32 columns

**Usage**

```
data(personality)
```

**Format**

This dataframe has recordings on the following 32 variables: distant, talkatv, carelss, hardwrk, anxious, agreeabl, tense, kind, opposng, relaxed, disorgn, outgoin, approvn, shy, discipl, harsh, persevr, friendl, worryin, respnsi, contrar, sociabl, lazy, coopera, quiet, organiz, criticl, lax, laidbck, withdrw, givinup, easygon

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**References**

Origin unclear

**Examples**

```
data(personality)
str(personality)
```

---

data\_cad      *Coronary artery disease data*

---

**Description**

A cross classified table with observational data from a Danish heart clinic. The response variable is CAD (coronary artery disease, some times called heart attack).

**Usage**

```
data(cad1)
```



**Format**

A data frame with 236 observations on the following 14 variables.

Sex Sex; a factor with levels Female Male

AngPec Angina pectoris (chest pain attacks); a factor with levels Atypical None Typical

AMI Acute myocardial infarct; a factor with levels Definite NotCertain

QWave A reading from an electrocardiogram; a factor with levels No Yes; Yes means pathological and is a sign of previous myocardial infarction.

QWavecode a factor with levels Nonusable Usable. An assesment of whether QWave is reliable.

STcode a factor with levels Nonusable Usable. An assesment of whether STchange is reliable.

STchange A reading from an electrocardiogram; a factor with levels No Yes. An STchange indicates a blockage of the coronary artery.

SuffHeartf Sufficient heart frequency; a factor with levels No, Yes

Hypertrophi a factor with levels No, Yes. Hypertrophy refers to an increased size of the heart muscle due to exercise.

Hyperchol a factor with levels No Yes. Hypercholesterolemia, also called high cholesterol, is the presence of high levels of cholesterol in the blood.

Smoker Is the patient a smoker; a factor with levels No, Yes.

Inherit Hereditary predispositions for CAD; a factor with levels No, Yes.

Heartfail Previous heart failures; a factor with levels No Yes

CAD Coronary Artery Disease; a factor with levels No Yes. CAD refers to a reduction of blood flow to the heart muscle (commonly known as a heart attack). The diagnosis made from biopsies.

**Details**

Notice that data are collected at a heart clinic, so data do not represent the population, but are conditional on patients having ended up at the clinic.

\* cad1: Complete dataset, 236 cases.

\* cad2: Incomplete dataset, 67 cases. Information on (some of) the variables 'Hyperchol', 'Smoker' and 'Inherit' is missing.

**References**

Hansen, J. F. (1980). The clinical diagnosis of ichaeme heart disease du to coronary artery disease. Danish Medical Bulletin

Højsgaard, Søren and Thiesson, Bo (1995). BIFROST - Block recursive models Induced From Relevant knowledge, Observations and Statistical Techniques. Computational Statistics and Data Analysis, vol. 19, p. 155-175

**Examples**

```
data(cad1)
## maybe str(cad1) ; plot(cad1) ...
```

---

descStat	<i>Computing simple descriptive statistics of a numeric vector.</i>
----------	---

---

**Description**

Computing simple descriptive statistics of a numeric vector - not unlike what proc means of SAS does

**Usage**

```
descStat(x, na.rm = TRUE)
```

**Arguments**

x	A numeric vector
na.rm	Should missing values be removed

**Value**

A vector with named elements.

**Author(s)**

Gregor Gorjanc; gregor.gorjanc <at> bf.uni-lj.si

**See Also**

[summaryBy](#), [summary\\_by](#)

**Examples**

```
x <- c(1, 2, 3, 4, NA, NaN)
descStat(x)
```

---

dietox	<i>Growth curves of pigs in a 3x3 factorial experiment</i>
--------	--

---

**Description**

The dietox data frame has 861 rows and 7 columns.

**Usage**

```
dietox
```

**Format**

This data frame contains the following columns:

**Weight** Weight in Kg

**Feed** Cumulated feed intake in Kg

**Time** Time (in weeks) in the experiment

**Pig** Factor; id of each pig

**Evit** Factor; vitamin E dose; see 'details'.

**Cu** Factor, copper dose; see 'details'

**Start** Start weight in experiment, i.e. weight at week 1.

**Litter** Factor, id of litter of each pig

**Details**

Data contains weight of slaughter pigs measured weekly for 12 weeks. Data also contains the startweight (i.e. the weight at week 1). The treatments are 3 different levels of Evit = vitamin E (dose: 0, 100, 200 mg dl-alpha-tocopheryl acetat /kg feed) in combination with 3 different levels of Cu=copper (dose: 0, 35, 175 mg/kg feed) in the feed. The cumulated feed intake is also recorded. The pigs are littermates.

**Source**

Lauridsen, C., Højsgaard, S., Sørensen, M.T. C. (1999) Influence of Dietary Rapeseed Oli, Vitamin E, and Copper on Performance and Antioxidant and Oxidative Status of Pigs. *J. Anim. Sci.* 77:906-916

**Examples**

```
data(dietox)
head(dietox)
if (require(ggplot2)){
  qplot(Time, Weight, data=dietox, col=Pig) + geom_line() +
    theme(legend.position = "none") + facet_grid(Evit~Cu)
} else {
  coplot(Weight ~ Time | Evit * Cu, data=dietox)
}
```

---

 esticon

*Contrasts for lm, glm, lme, and geeglm objects*


---

## Description

Computes linear functions (i.e. weighted sums) of the estimated regression parameters. Can also test the hypothesis, that such a function is equal to a specific value.

## Usage

```
esticon(obj, L, beta0, conf.int = TRUE, level = 0.95, joint.test = FALSE, ...)

## S3 method for class 'esticon_class'
coef(object, ...)

## S3 method for class 'esticon_class'
summary(object, ...)

## S3 method for class 'esticon_class'
confint(object, parm, level = 0.95, ...)

## S3 method for class 'esticon_class'
vcov(object, ...)
```

## Arguments

obj	Regression object (of type lm, glm, lme, geeglm).
L	Matrix (or vector) specifying linear functions of the regression parameters (one linear function per row). The number of columns must match the number of fitted regression parameters in the model. See 'details' below.
beta0	A vector of numbers
conf.int	TRUE
level	The confidence level
joint.test	Logical value. If TRUE a 'joint' Wald test for the hypothesis $L\beta = \beta_0$ is made. Default is that the 'row-wise' tests are made, i.e. $(L\beta)_i = \beta_{0i}$ . If joint.test is TRUE, then no confidence interval etc. is calculated.
...	Additional arguments; currently not used.
object	An esticon_class object.
parm	a specification of which parameters are to be given confidence intervals, either a vector of numbers or a vector of names. If missing, all parameters are considered.

**Details**

Let the estimated parameters of the model be

$$\beta_1, \beta_2, \dots, \beta_p$$

A linear function of the estimates is of the form

$$l = \lambda_1\beta_1 + \lambda_2\beta_2 + \dots + \lambda_p\beta_p$$

where  $\lambda_1, \lambda_2, \dots, \lambda_p$  is specified by the user.

The esticon function calculates  $l$ , its standard error and by default also a 95 pct confidence interval. It is sometimes of interest to test the hypothesis  $H_0 : l = \beta_0$  for some value  $\beta_0$  given by the user. A test is provided for the hypothesis  $H_0 : l = 0$  but other values of  $\beta_0$  can be specified.

In general, one can specify  $r$  such linear functions at one time by specifying  $L$  to be an  $r \times p$  matrix where each row consists of  $p$  numbers  $\lambda_1, \lambda_2, \dots, \lambda_p$ . Default is then that  $\beta_0$  is a  $p$  vector of 0s but other values can be given.

It is possible to test simultaneously that all specified linear functions are equal to the corresponding values in  $\beta_0$ .

For computing contrasts among levels of a single factor, 'contrast.lm' may be more convenient.

**Value**

Returns a matrix with one row per linear function. Columns contain estimated coefficients, standard errors, t values, degrees of freedom, two-sided p-values, and the lower and upper endpoints of the 1-alpha confidence intervals.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**Examples**

```
data(iris)
lm1 <- lm(Sepal.Length ~ Sepal.Width + Species + Sepal.Width : Species, data=iris)
## Note that the setosa parameters are set to zero
coef(lm1)

## Estimate the intercept for versicolor
lambda1 <- c(1, 0, 1, 0, 0, 0)
esticon(lm1, L=lambda1)

## Estimate the difference between versicolor and virgica intercept
## and test if the difference is 1
lambda2 <- c(0, 1, -1, 0, 0, 0)
esticon(lm1, L=lambda2, beta0=1)

## Do both estimates at one time
esticon(lm1, L=rbind(lambda1, lambda2), beta0=c(0, 1))
```

```
## Make a combined test for that the difference between versicolor and virgica intercept
## and difference between versicolor and virginica slope is zero:
lambda3 <- c(0, 0, 0, 0, 1, -1)
esticon(lm1, L=rbind(lambda2, lambda3), joint.test=TRUE)

# Example using esticon on coxph objects (thanks to Alessandro A. Leidi).
# Using dataset 'veteran' in the survival package
# from the Veterans' Administration Lung Cancer study

if (require(survival)){
  data(veteran)
  sapply(veteran, class)
  levels(veteran$celltype)
  attach(veteran)
  veteran.s <- Surv(time, status)
  coxmod <- coxph(veteran.s ~ age + celltype + trt, method='breslow')
  summary(coxmod)

  # compare a subject 50 years old with celltype 1
  # to a subject 70 years old with celltype 2
  # both subjects on the same treatment
  AvB <- c(-20, -1, 0, 0, 0)

  # compare a subject 40 years old with celltype 2 on treat=0
  # to a subject 35 years old with celltype 3 on treat=1
  CvB <- c(5, 1, -1, 0, -1)

  est <- esticon(coxmod, L=rbind(AvB, CvB))
  est
  ##exp(est[, c(2, 7, 8)])
}
```

---

expr\_to\_fun

*Convert expression into function object.*

---

### **Description**

Convert expression into function object.

### **Usage**

```
expr_to_fun(e)
```

### **Arguments**

e                    expression

**Examples**

```
ee <- expression(b1 + (b0 - b1)*exp(-k*x) + b2*x)
ff <- expr_to_fun(ee)
```

---

fatacid	<i>Fish oil in pig food</i>
---------	-----------------------------

---

**Description**

Fish oil in pig food

**Usage**

fatacid

**Format**

A dataframe.

**Details**

A fish oil fatty acid X14 has been added in different concentrations to the food for pigs in a study. Interest is in studying how much of the fatty acid can be found in the tissue. The concentrations of x14 in the food are  $\text{verb+dose}=\{0.0, 4.4, 6.2, 9.3\}$ .

The pigs are fed with this food until their weight is 60 kg. From thereof and until they are slaughtered at 100kg, their food does not contain the fish oil. At 60kg (sample=1) and 100kg (sample=2) muscle biopsies are made and the concentration of x14 is determined. Measurements on the same pig are correlated, and pigs are additionally related through litters.

**References**

Data courtesy of Charlotte Lauridsen, Department of Animal Science, Aarhus University, Denmark.

---

fev	<i>Forced expiratory volume in children</i>
-----	---

---

**Description**

Dataset to examine if respiratory function in children was influenced by smoking.

**Usage**

fev

**Format**

A data frame with 654 observations on the following 5 variables.

Age Age in years.

FEV Forced expiratory volume in liters per second.

Ht Height in centimeters.

Gender Gender.

Smoke Smoking status.

**References**

I. Tager and S. Weiss and B. Rosner and F. Speizer (1979). Effect of Parental Cigarette Smoking on the Pulmonary Function of Children. *American Journal of Epidemiology*. 110:15-26

**Examples**

```
data(fev)
summary(fev)
```

---

firstlastobs	<i>Locate the index of the first/last unique value</i>
--------------	--

---

**Description**

Locate the index of the first/last unique value in i) a vector or of a variable in a data frame.

**Usage**

```
lastobs(x, ...)

firstobs(x, ...)

## Default S3 method:
lastobs(x, ...)

## Default S3 method:
firstobs(x, ...)

## S3 method for class 'formula'
lastobs(formula, data = parent.frame(), ...)

## S3 method for class 'formula'
firstobs(formula, data = parent.frame(), ...)
```



**Arguments**

x	A vector
...	Currently not used
formula	A formula (only the first term is used, see 'details').
data	A data frame

**Details**

If writing  $\sim a + b + c$  as formula, then only a is considered.

**Value**

A vector.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**Examples**

```
x <- c(rep(1, 5), rep(2, 3), rep(3, 7), rep(1, 4))
firstobs(x)
lastobs(x)
data(dietox)
firstobs(~Pig, data=dietox)
lastobs(~Pig, data=dietox)
```

---

haldCement

*Heat development in cement under hardening.*

---

**Description**

Heat development in cement under hardening related to the chemical composition.

**Usage**

```
haldCement
```

**Format**

A data frame with 13 observations on the following 5 variables.

x1 Percentage (weight) of  $[3\text{CaO}][\text{Al}_2\text{O}_3]$

x2 Percentage (weight) of  $[3\text{CaO}][\text{SiO}_2]$

x3 Percentage (weight) of  $[4\text{CaO}][\text{Al}_2\text{O}_3][\text{FeO}_3]$

x4 Percentage (weight) of  $[2\text{CaO}][\text{SiO}_2]$

y Heat development measured in calories per gram cement after 180 days

**References**

Anders Hald (1949); Statistiske Metoder; Akademisk Forlag (in Danish), page 509.

**Examples**

```
data(haldCement)

if( interactive() ){
  pairs( haldCement )
}
m <- lm( y ~ x1 + x2 + x3 + x4, data=haldCement )
summary( m )

# Notice: The model explains practically all variation in data;
# yet none of the explanatory variables appear to be statistically
# significant.
```

---

interaction-plot	<i>Two-way interaction plot</i>
------------------	---------------------------------

---

**Description**

Plots the mean of the response for two-way combinations of factors, thereby illustrating possible interactions.

**Usage**

```
interaction_plot(.data, .formula, interval = "conf.int")
```

**Arguments**

.data	A data frame
.formula	A formula of the form 'y ~ x1 + x2'
interval	Either 'conf.int', 'boxplot' or 'none'

**Note**

This is a recent addition to the package and is subject to change.

**Examples**

```
ToothGrowth %>% interaction_plot(len ~ dose + supp)
ToothGrowth %>% interaction_plot(len ~ dose + supp, interval="conf.int")
ToothGrowth %>% interaction_plot(len ~ dose + supp, interval="boxplot")
ToothGrowth %>% interaction_plot(len ~ dose + supp, interval="none")
```

---

is-estimable	<i>Determines if contrasts are estimable.</i>
--------------	---

---

**Description**

Determines if contrasts are estimable, that is, if the contrasts can be written as a linear function of the data.

**Usage**

```
is_estimable(K, null.basis)
```

**Arguments**

K	A matrix.
null.basis	A basis for a null space (can be found with <code>null_basis()</code> ).

**Details**

Consider the setting  $E(Y) = Xb$ . A linear function of  $b$ , say  $l'b$  is estimable if and only if there exists an  $r$  such that  $r'X = l'$  or equivalently  $l = X'r$ . Hence  $l$  must be in the column space of  $X'$ , i.e. in the orthogonal complement of the null space of  $X$ . Hence, with a basis  $B$  for the null space, `is_estimable()` checks if each row  $l$  of the matrix  $K$  is perpendicular to each column basis vector in  $B$ .

**Value**

A logical vector.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**References**

[http://web.mit.edu/18.06/www/Essays/newpaper\\_ver3.pdf](http://web.mit.edu/18.06/www/Essays/newpaper_ver3.pdf)

**See Also**

[null\\_basis](#)

**Examples**

```
## TO BE WRITTEN
```

---

linest                      *Compute linear estimates*

---

### Description

Compute linear estimates, i.e. 'L linear estimates is population means (also known as LSMEANS).

### Usage

```
linest(object, L = NULL, ...)

## S3 method for class 'linest_class'
confint(object, parm, level = 0.95, ...)

## S3 method for class 'linest_class'
coef(object, ...)

## S3 method for class 'linest_class'
summary(object, ...)
```

### Arguments

object	Model object
L	Either NULL or a matrix with p columns where p is the number of parameters in the systematic effects in the model. If NULL then L is taken to be the p times p identity matrix
...	Additional arguments; currently not used.
parm	Specification of the parameters estimates for which confidence intervals are to be calculated.
level	The level of the (asymptotic) confidence interval.
confint	Should confidence interval appear in output.

### Value

A dataframe with results from computing the contrasts.

### Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

### See Also

[LSmeans](#), [LE\\_matrix](#)

**Examples**

```
## Make balanced dataset
dat.bal <- expand.grid(list(AA=factor(1:2), BB=factor(1:3), CC=factor(1:3)))
dat.bal$y <- rnorm(nrow(dat.bal))

## Make unbalanced dataset
# 'BB' is nested within 'CC' so BB=1 is only found when CC=1
# and BB=2,3 are found in each CC=2,3,4
dat.nst <- dat.bal
dat.nst$CC <-factor(c(1,1,2,2,2,2,1,1,3,3,3,3,1,1,4,4,4,4))

mod.bal <- lm(y ~ AA + BB * CC, data=dat.bal)
mod.nst <- lm(y ~ AA + BB : CC, data=dat.nst)

L <- LE_matrix(mod.nst, effect=c("BB", "CC"))
linest( mod.nst, L )
```

---

linest-get

*Auxillary functions for computing lsmeans, contrasts etc*


---

**Description**

Auxillary functions for computing lsmeans, contrasts etc.

**Usage**

```
get_xlevels(obj)

## Default S3 method:
get_xlevels(obj)

## S3 method for class 'mer'
get_xlevels(obj)

## S3 method for class 'merMod'
get_xlevels(obj)

get_contrasts(obj)

## Default S3 method:
get_contrasts(obj)

## S3 method for class 'merMod'
get_contrasts(obj)

set_xlevels(xlev, at)
```

```

get_vartypes(obj)

set_covariate_val(xlev, covariateVal)

get_X(obj, newdata, at = NULL)

## Default S3 method:
get_X(obj, newdata, at = NULL)

## S3 method for class 'merMod'
get_X(obj, newdata, at = NULL)

```

### Arguments

obj	An R object
xlev	FIXME: to be described
at	FIXME: to be described
covariateVal	FIXME: to be described
newdata	FIXME: to be described

---

linest-matrix	<i>Linear estimates matrix</i>
---------------	--------------------------------

---

### Description

Generate matrix specifying linear estimate.

### Usage

```

LE_matrix(object, effect = NULL, at = NULL)

## Default S3 method:
LE_matrix(object, effect = NULL, at = NULL)

aggregate_linest_list(linest_list)

get_linest_list(object, effect = NULL, at = NULL)

```

### Arguments

object	Model object
effect	A vector of variables. For each configuration of these the estimate will be calculated.

`at` Either NULL, a list or a dataframe. 1) If a list, then the list must consist of covariates (including levels of some factors) to be used in the calculations. 2) If a dataframe, the dataframe is split rowwise and the function is invoked on each row.

`linest_list` Linear estimate list (as generated by `get_linest_list`).

## Details

Check this

## See Also

[LSmeans](#), [linest](#)

## Examples

```
## Two way anova:
data(warpbreaks)

## An additive model
m0 <- lm(breaks ~ wool + tension, data=warpbreaks)

## Estimate mean for each wool type, for tension="M":
K <- LE_matrix(m0, at=list(wool=c("A", "B"), tension="M"))
K

## Vanilla computation:
K %*% coef(m0)

## Alternative; also providing standard errors etc:
linest(m0, K)
esticon(m0, K)

## Estimate mean for each wool type when averaging over tension;
# two ways of doing this
K <- LE_matrix(m0, at=list(wool=c("A", "B")))
K
K <- LE_matrix(m0, effect="wool")
K
linest(m0, K)

## The linear estimate is sometimes called to "least squares mean"
# (LSmeans) or population means.
# Same as
LSmeans(m0, effect="wool")

## Without mentioning 'effect' or 'at' an average across all
#predictors are calculated:
K <- LE_matrix(m0)
K
```

```

linest(m0, K)

## Because the design is balanced (9 observations per combination
## of wool and tension) this is the same as computing the average. If
## the design is not balanced, the two quantities are in general not
## the same.
mean(warpbreaks$breaks)

## Same as
LSmeans(m0)

## An interaction model
m1 <- lm(breaks ~ wool * tension, data=warpbreaks)

K <- LE_matrix(m1, at=list(wool=c("A", "B"), tension="M"))
K
linest(m1, K)
K <- LE_matrix(m1, at=list(wool=c("A", "B")))
K
linest(m1, K)
K <- LE_matrix(m1, effect="wool")
K
linest(m1, K)
LSmeans(m1, effect="wool")

K <- LE_matrix(m1)
K
linest(m1, K)
LSmeans(m1)

```

---

ls-means

---

*Compute LS-means (aka population means or marginal means)*


---

### Description

LS-means (least squares means, also known as population means and as marginal means) for a range of model types.

### Usage

```

LSmeans(object, effect = NULL, at = NULL, level = 0.95, ...)

## Default S3 method:
LSmeans(object, effect = NULL, at = NULL, level = 0.95, ...)

## S3 method for class 'lmerMod'
LSmeans(object, effect = NULL, at = NULL, level = 0.95, adjust.df = TRUE, ...)

popMeans(object, effect = NULL, at = NULL, level = 0.95, ...)

```



```
## Default S3 method:
popMeans(object, effect = NULL, at = NULL, level = 0.95, ...)

## S3 method for class 'lmerMod'
popMeans(object, effect = NULL, at = NULL, level = 0.95, adjust.df = TRUE, ...)
```

### Arguments

<code>object</code>	Model object
<code>effect</code>	A vector of variables. For each configuration of these the estimate will be calculated.
<code>at</code>	A list of values of covariates (including levels of some factors) to be used in the calculations
<code>level</code>	The level of the (asymptotic) confidence interval.
<code>...</code>	Additional arguments; currently not used.
<code>adjust.df</code>	Should denominator degrees of freedom be adjusted?

### Details

There are restrictions on the formulas allowed in the model object. For example having  $y \sim \log(x)$  will cause an error. Instead one must define the variable  $\logx = \log(x)$  and do  $y \sim \logx$ .

### Value

A dataframe with results from computing the contrasts.

### Warning

Notice that `LSmeans` and `LE_matrix` fails if the model formula contains an offset (as one would have in connection with e.g. Poisson regression). It is on the todo-list to fix this

### Note

`LSmeans` and `popMeans` are synonymous. Some of the code has been inspired by the `lsmeans` package.

### Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

### See Also

[LE\\_matrix](#), [linest](#)

**Examples**

```

## Two way anova:

data(warpbreaks)

m0 <- lm(breaks ~ wool + tension, data=warpbreaks)
m1 <- lm(breaks ~ wool * tension, data=warpbreaks)
LSmeans(m0)
LSmeans(m1)

## same as:
K <- LE_matrix(m0);K
linest(m0, K)
K <- LE_matrix(m1);K
linest(m1, K)

LE_matrix(m0, effect="wool")
LSmeans(m0, effect="wool")

LE_matrix(m1, effect="wool")
LSmeans(m1, effect="wool")

LE_matrix(m0, effect=c("wool", "tension"))
LSmeans(m0, effect=c("wool", "tension"))

LE_matrix(m1, effect=c("wool", "tension"))
LSmeans(m1, effect=c("wool", "tension"))

## Regression; two parallel regression lines:

data(Puromycin)

m0 <- lm(rate ~ state + log(conc), data=Puromycin)
## Can not use LSmeans / LE_matrix here because of
## the log-transformation. Instead we must do:
Puromycin$lconc <- log( Puromycin$conc )
m1 <- lm(rate ~ state + lconc, data=Puromycin)

LE_matrix(m1)
LSmeans(m1)

LE_matrix(m1, effect="state")
LSmeans(m1, effect="state")

LE_matrix(m1, effect="state", at=list(lconc=3))
LSmeans(m1, effect="state", at=list(lconc=3))

## Non estimable contrasts

## ## Make balanced dataset

```

```

dat.bal <- expand.grid(list(AA=factor(1:2), BB=factor(1:3),
                          CC=factor(1:3)))
dat.bal$y <- rnorm(nrow(dat.bal))

## ## Make unbalanced dataset
#      'BB' is nested within 'CC' so BB=1 is only found when CC=1
#      and BB=2,3 are found in each CC=2,3,4
dat.nst <- dat.bal
dat.nst$CC <- factor(c(1, 1, 2, 2, 2, 2, 1, 1, 3, 3,
                      3, 3, 1, 1, 4, 4, 4, 4))

mod.bal <- lm(y ~ AA + BB * CC, data=dat.bal)
mod.nst <- lm(y ~ AA + BB : CC, data=dat.nst)

LSmeans(mod.bal, effect=c("BB", "CC"))
LSmeans(mod.nst, effect=c("BB", "CC"))
LSmeans(mod.nst, at=list(BB=1, CC=1))

LSmeans(mod.nst, at=list(BB=1, CC=2))
## Above: NA's are correct; not an estimable function

if( require( lme4 )){
  warp.mm <- lmer(breaks ~ -1 + tension + (1|wool), data=warpbreaks)
  LSmeans(warp.mm, effect="tension")
  class(warp.mm)
  fixef(warp.mm)
  coef(summary(warp.mm))
  vcov(warp.mm)
  if (require(pbkrtest))
    vcovAdj(warp.mm)
}

LSmeans(warp.mm, effect="tension")

```

---

mb\_summary

*Fast summary of microbenchmark object*


---

## Description

Fast summary of microbenchmark object. The default summary method from the microbenchmark package is fairly slow in producing a summary (due to a call to a function from the multcomp package.)

## Usage

```
mb_summary(object, unit, add.unit = TRUE, ...)
```

```
summary_mb(object, unit, add.unit = TRUE, ...)
```

**Arguments**

object	A microbenchmark object
unit	The time unit to be used
add.unit	Should time unit be added as column to resulting dataframe.
...	Additional arguments; currently not used.

---

milkman	<i>Milk yield data for manually milked cows.</i>
---------	--

---

**Description**

Milk yield data for cows milked manually twice a day (morning and evening).

**Usage**

```
milkman
```

**Format**

A data frame with 161836 observations on the following 12 variables.

cowno a numeric vector; cow identification

lactno a numeric vector; lactation number

ampm a numeric vector; milking time: 1: morning; 2: evening

dfc a numeric vector; days from calving

my a numeric vector; milk yield (kg)

fatpct a numeric vector; fat percentage

protpct a numeric vector; protein percentage

lactpct a numeric vector; lactose percentage

scc a numeric vector; somatic cell counts

race a factor with levels RDM Holstein Jersey

ecmy a numeric vector; energy corrected milk

cowlact Combination of cowno and lactno; necessary because the same cow may appear more than once in the dataset (in different lactations)

**Details**

There are data for 222 cows. Some cows appear more than once in the dataset (in different lactations) and there are 288 different lactations.

## References

Friggens, N. C.; Ridder, C. and Løvendahl, P. (2007). On the Use of Milk Composition Measures to Predict the Energy Balance of Dairy Cows. *J. Dairy Sci.* 90:5453–5467 doi:10.3168/jds.2006-821.

This study was part of the Biosens project used data from the “Mælkekoens energibalance og mobilisering” project; both were funded by the Danish Ministry of Food, Agriculture and Fisheries and the Danish Cattle Association.

## Examples

```
data(milkman)
```

---

NIRmilk

*NIRmilk*

---

## Description

Near infra red light (NIR) measurements are made at 152 wavelengths on 17 milk samples. While milk runs through a glass tube, infra red light is sent through the tube and the amount of light passing through the tube is measured at different wavelengths. Each milk sample was additionally analysed for fat, lactose, protein and drymatter.

## Usage

```
NIRmilk
```

## Format

This data frame contains 18 rows and 158 columns. The first column is the sample number. The columns X<sub>www</sub> contains the infra red light amount at wavelength www. The response variables are fat, protein, lactose and dm (drymatter).

## Examples

```
data(NIRmilk)
```

---

null-basis	<i>Finds the basis of the (right) null space.</i>
------------	---

---

### Description

Finds the basis of the (right) null space of a matrix, a vector (a 1-column matrix) or a model object for which a model matrix can be extracted. I.e. finds basis for the (right) null space  $x : Mx = 0$ .

### Usage

```
null_basis(object)
```

### Arguments

object	A matrix, a vector (a 1-column matrix) or a model object for which a model matrix can be extracted (using <code>model.matrix</code> ).
--------	--

### Value

A matrix (possibly with zero columns if the null space consists only of the zero vector).

### Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

### See Also

[Null](#)

### Examples

```
M <- matrix(c(1,1,1,1,1,1,0,0,0,0,1,1), nrow=4)
null_basis(M)
MASS::Null(t(M))

M <- c(1,1,1,1)
null_basis(M)
MASS::Null(t(M))

m0 <- lm(breaks ~ wool + tension, data=warpbreaks)
null_basis(m0)
MASS::Null(t(model.matrix(m0)))

## Make balanced dataset
dat.bal <- expand.grid(list(A=factor(1:2), B=factor(1:3), C=factor(1:3)))
dat.bal$y <- rnorm(nrow(dat.bal))

## Make unbalanced dataset: 'B' is nested within 'C' so B=1 is only
```

```
## found when C=1 and B=2,3 are found in each C=2,3,4
dat.nst <- dat.bal
dat.nst$C <-factor(c(1,1,2,2,2,2,1,1,3,3,3,3,1,1,4,4,4,4))
xtabs(y ~ C+B+A , data=dat.nst)

mod.bal <- lm(y ~ A + B*C, data=dat.bal)
mod.nst <- lm(y ~ A + B*C, data=dat.nst)

null_basis( mod.bal )
null_basis( mod.nst )

null_basis( model.matrix(mod.bal) )
null_basis( model.matrix(mod.nst) )

MASS::Null( t(model.matrix(mod.bal)) )
MASS::Null( t(model.matrix(mod.nst)) )
```

---

parseGroupFormula      *Extract components from a formula with "conditioning bar"*

---

## Description

Extract components from a formula with the form  $y \sim x_1 + \dots + x_n \mid g_1 + \dots + g_m$

## Usage

```
parseGroupFormula(form)
```

## Arguments

form                    A formula of the form  $y \sim x_1 + \dots + x_n \mid g_1 + \dots + g_m$

## Value

If the formula is  $y \sim x_1 + x_2 \mid g_1 + g_2$  the result is

```
model                    y ~ x1 + x2
groups                   g1 + g2
groupFormula             ~ g1 + g2
```

## Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

## Examples

```
gf <- parseGroupFormula(y ~ x1 + x2 | g1 + g2)
gf
```

---

potatoes	<i>Weight and size of 20 potatoes</i>
----------	---------------------------------------

---

**Description**

Weight and size of 20 potatoes. Weight in grams; size in milimeter. There are two sizes: length is the longest length and width is the shortest length across a potato. #'

**Usage**

```
potatoes
```

**Format**

A data frame with 20 observations on the following 3 variables.

weight a numeric vector

length a numeric vector

width a numeric vector

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**Source**

My own garden; autumn 2015.

**Examples**

```
data(potatoes)
plot(potatoes)
```

---

prostate	<i>Prostate Tumor Gene Expression Dataset</i>
----------	---

---

**Description**

This is the Prostate Tumor Gene Expression dataset used in Chung and Keles (2010).

**Usage**

```
data(prostate)
```



**Format**

A list with two components:

**x** Gene expression data. A matrix with 102 rows and 6033 columns.

**y** Class index. A vector with 102 elements.

**Details**

The prostate dataset consists of 52 prostate tumor and 50 normal samples. Normal and tumor classes are coded in 0 and 1, respectively, in **y** vector. Matrix **x** is gene expression data and arrays were normalized, log transformed, and standardized to zero mean and unit variance across genes as described in Dettling (2004) and Dettling and Beuhlmann (2002). See Chung and Keles (2010) for more details.

**Source**

Singh D, Febbo P, Ross K, Jackson D, Manola J, Ladd C, Tamayo P, Renshaw A, D'Amico A, Richie J, Lander E, Loda M, Kantoff P, Golub T, and Sellers W (2002), "Gene expression correlates of clinical prostate cancer behavior", *Cancer Cell*, Vol. 1, pp. 203–209.

**References**

Chung D and Keles S (2010), "Sparse partial least squares classification for high dimensional data", *Statistical Applications in Genetics and Molecular Biology*, Vol. 9, Article 17.

Dettling M (2004), "BagBoosting for tumor classification with gene expression data", *Bioinformatics*, Vol. 20, pp. 3583–3593.

Dettling M and Beuhlmann P (2002), "Supervised clustering of genes", *Genome Biology*, Vol. 3, pp. research0069.1–0069.15.

**Examples**

```
data(prostate)
prostate$x[1:5,1:5]
prostate$y
```

---

recodeVar

*Recode values of a vector*

---

**Description**

Recodes a vector with values, say 1,2 to a variable with values, say 'a', 'b'

**Usage**

```
recodeVar(x, src, tgt, default = NULL, keep.na = TRUE)
```

**Arguments**

x	A vector; the variable to be recoded
src	The source values: a subset of the present values of x
tgt	The target values: the corresponding new values of x
default	Default target value for those values of x not listed in 'src'. When default=NULL, values of x which are not given in 'src' will be kept in the output.
keep.na	If TRUE then NA's in x will be retained in the output

**Value**

A vector

**Warning**

Care should be taken if x is a factor. A safe approach may be to convert x to a character vector using `as.character`.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[cut](#), [factor](#), [recodeVar](#)

**Examples**

```
x <- c("dec", "jan", "feb", "mar", "apr", "may")
src1 <- list(c("dec", "jan", "feb"), c("mar", "apr", "may"))
tgt1 <- list("winter", "spring")
recodeVar(x, src=src1, tgt=tgt1)
#[1] "winter" "winter" "winter" "spring" "spring" "spring"

x <- c(rep(1:3, 3))
#[1] 1 2 3 1 2 3 1 2 3

## Simple usage:
recodeVar(x, src=c(1, 2), tgt=c("A", "B"))
#[1] "A" "B" NA "A" "B" NA "A" "B" NA

## Here we need to use lists
recodeVar(x, src=list(c(1, 2)), tgt=list("A"))
#[1] "A" "A" NA "A" "A" NA "A" "A" NA
recodeVar(x, src=list(c(1, 2)), tgt=list("A"), default="L")
#[1] "A" "A" "L" "A" "A" "L" "A" "A" "L"
recodeVar(x, src=list(c(1, 2), 3), tgt=list("A", "B"), default="L")
#[1] "A" "A" "B" "A" "A" "B" "A" "A" "B"

## Dealing with NA's in x
```

```

x<-c(NA,rep(1:3, 3),NA)
#[1] NA 1 2 3 1 2 3 1 2 3 NA
recodeVar(x, src=list(c(1, 2)), tgt=list("A"))
#[1] NA "A" "A" NA "A" "A" NA "A" "A" NA NA
recodeVar(x, src=list(c(1, 2)), tgt=list("A"), default="L")
#[1] NA "A" "A" "L" "A" "A" "L" "A" "A" "L" NA
recodeVar(x, src=list(c(1, 2)), tgt=list("A"), default="L", keep.na=FALSE)
#[1] "L" "A" "A" "L" "A" "A" "L" "A" "A" "L" "L"

x <- c("no", "yes", "not registered", "no", "yes", "no answer")
recodeVar(x, src = c("no", "yes"), tgt = c("0", "1"), default = NA)

```

---

recover_pca_data	<i>Recover data from principal component analysis</i>
------------------	---

---

## Description

Recover data from principal component analysis based on the first (typically few) components.

## Usage

```
recover_pca_data(object, comp = 1)
```

## Arguments

object	An object of class 'prcomp'.
comp	The number of components to be used. Must be smaller than the number of variables.

## Value

A dataframe

## Examples

```

crime <- doBy::crimeRate
rownames(crime) <- crime$State
crime$State <- NULL
crime <- doBy::crimeRate
rownames(crime) <- crime$State
crime$State <- NULL

o <- order(apply(scale(crime), 1, sum))
dat <- crime[o,]
head(dat)
tail(dat)

```

```
matplot(scale(dat), type="l")

pc1 <- prcomp(dat, scale. = TRUE)
summary(pc1)
rec2 <- recover_pca_data(pc1, 2)

pairs(rec2)

par(mfrow=c(1,2))
matplot(scale(dat), type="l")
matplot(scale(rec2), type="l")

j <- merge(dat, rec2, by=0)
pairs(j[,-1])
```

---

**renameCol***Rename columns in a matrix or a dataframe.*

---

### Description

Rename columns in a matrix or a dataframe.

### Usage

```
renameCol(indata, src, tgt)
```

### Arguments

indata	A dataframe or a matrix
src	Source: Vector of names of columns in 'indata' to be renamed. Can also be a vector of column numbers.
tgt	Target: Vector with corresponding new names in the output.

### Value

A dataframe if 'indata' is a dataframe; a matrix in 'indata' is a matrix.

### Author(s)

Søren Højsgaard, <sorenh@math.aau.dk>

**Examples**

```

renameCol(CO2, 1:2, c("kk", "ll"))
renameCol(CO2, c("Plant", "Type"), c("kk", "ll"))

# These fail - as they should:
# renameCol(CO2, c("Plant", "Type", "conc"), c("kk", "ll"))
# renameCol(CO2, c("Plant", "Type", "Plant"), c("kk", "ll"))

```

---

restrict_fun	<i>restrict</i>
--------------	-----------------

---

**Description**

Restrict a functions domain by fixing certain arguments of a function call.

**Usage**

```

restrict_fun(fun, args, method = "env")

restrict_fun_sub(fun, args, envir = parent.frame())

restrict_fun_env(fun, args)

get_restrictions(object)

get_fun(object)

```

**Arguments**

fun	Function to be restricted
args	List of the form name=value
method	Either "env" (for environment; the default, using an auxillary argument for storing restricted values) or "sub" (for substitute; based on substituting fixed values into the function).
envir	Environment
object	An object from restrict_fun (a scaffold object).

**Details**

'restrict\_fun' is a wrapper for calling 'restrict\_fun\_env' (default) or 'restrict\_fun\_sub'.

**Value**

A new function: The input function 'fun' but with certain arguments fixed at specific values.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk> based on code adapted from the curry package.

**Examples**

```
f1 <- function(x, y){x + y}
f1_ <- restrict_fun(f1, list(y=10))
f1_
f1_(x=1)
get_restrictions(f1_)
get_fun(f1_)

f2 <- function(x){
  x <- x + 2
  x
}
f2_ <- restrict_fun(f2, list(x=1))
f2_()

# Notice that this is absurd, because arguments are modified in the
# body
restrict_fun(f2, list(x=10), method="sub")

# A safe(r) alternative is:
f3 <- function(x){
  x_ <- x + 2
  x_
}
restrict_fun(f3, list(x=10), method="sub")

rnorm10 <- restrict_fun(rnorm, list(n=10))
rnorm(10)
```

---

scale2

*Scale a dataframe or matrix*

---

**Description**

Similar to 'base::scale' but scales / centers only numeric values in data.

**Usage**

```
scale2(x, center = TRUE, scale = TRUE)
```

**Arguments**

x	dataframe or matrix
center	Logical, should data be centered.
scale	Logical, should data be scaled.

**Value**

An object of same class as 'x'

**Examples**

```
scale2(iris)
```

---

split_byrow_bycol	<i>Split matrix or dataframe into list</i>
-------------------	--

---

**Description**

Split matrix or dataframe into list by columns or by rows

**Usage**

```
split_bycol(x)
```

```
split_byrow(x)
```

**Arguments**

x	Matrix or dataframe
---	---------------------

---

sub_seq	<i>Find sub-sequences of identical elements in a vector.</i>
---------	--

---

**Description**

Find sub-sequences of identical elements in a vector.

**Usage**

```
subSeq(x, item = NULL)
```

```
sub_seq(x, item = NULL)
```

```
is_grouped(x)
```

```
rle2(x)
```

**Arguments**

x	An atomic vector or a factor.
item	Optionally a specific value to look for in 'x'.

**Details**

- \* 'sub\_seq' is synonymous with 'subSeq'
- \* 'rle2' is identical to 'rle' (from base) but 'rle2' works on factors as input (a factor is coerced to character).
- \* 'is\_grouped' checks if the values in 'x' are clustered into the smallest number of clusters.

**Value**

A dataframe.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[rle](#)

**Examples**

```
x <- c(1, 1, 1, 0, 0, 1, 1, 1, 2, 2, 2, 1, 2, 2, 2, 3)
(ans <- subSeq(x))
ans$value
# Notice: Same results below
subSeq(x, item=1)
subSeq(x, item="1")

xc <- as.character(x)
(ans<-subSeq(xc))
ans$value
# Notice: Same results below
subSeq(xc, item="1")
subSeq(xc, item=1)

is_grouped(x)
is_grouped(sort(x))
is_grouped(xc)
is_grouped(sort(xc))
```

---

taylor

*Taylor expansion (one dimension)*

---

**Description**

Returns Taylor polynomial approximating a function  $fn(x)$



**Usage**

```
taylor(fn, x0, ord = 1)
```

**Arguments**

fn	A function of one variable and that variable must be named 'x'.
x0	The point in which to to the Taylor expansion.
ord	The order of the Taylor expansion.

**Value**

function.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**Examples**

```
fn <- function(x) log(x)
ord <- 2
x0 <- 2

xv <- seq(.2, 5, .1)
plot(xv, fn(xv), type="l")
lines(xv, taylor(fn, x0=x0, ord=ord)(xv), lty=2)
abline(v=x0)

fn <- function(x)sin(x)
ord <- 4
x0 <- 0
xv <- seq(-2*pi, 2*pi, 0.1)
plot(xv, fn(xv), type="l")
lines(xv, taylor(fn, x0=x0, ord=ord)(xv), lty=2)
abline(v=x0)
```

---

tidy-esticon

*Tidy an esticon object*

---

**Description**

Tidy summarizes information about the components of the object.

**Usage**

```
## S3 method for class 'esticon_class'
tidy(x, conf.int = FALSE, conf.level = 0.95, ...)
```

**Arguments**

x	A 'esticon_class' object (produced by esticon methods).
conf.int	Should confidence intervals be added.
conf.level	Desired confidence level.
...	Additional arguments; currently not used.

---

tidy-linest	<i>Tidy a linest object</i>
-------------	-----------------------------

---

**Description**

Tidy summarizes information about the components of the object.

**Usage**

```
## S3 method for class 'linest_class'
tidy(x, conf.int = FALSE, conf.level = 0.95, ...)
```

**Arguments**

x	A 'linest_class' object (produced by linest methods).
conf.int	Should confidence intervals be added.
conf.level	Desired confidence level.
...	Additional arguments; currently not used.

---

timeSinceEvent	<i>Calculate "time since event" in a vector.</i>
----------------	--

---

**Description**

Calculate "time since event" in a vector.

**Usage**

```
timeSinceEvent(yvar, tvar = seq_along(yvar))
```

**Arguments**

yvar	A numerical or logical vector specifying the events
tvar	An optional vector specifying time

**Details**

Events are coded as 1 in numeric vector (and non-events are coded with values different from 1). `timeSinceEvent` will give the time since event (with and without sign). In a logical vector, events are coded as TRUE and all non-events as FALSE.

**Value**

A dataframe with columns 'yvar', 'tvar', 'abs.tse' (absolute time since nearest event), 'sign.tse' (signed time since nearest event) and 'run' (indicator of the time window around each event).

**Note**

NA's in yvar are converted to zeros.

**Author(s)**

Søren Højsgaard, <sorenh@math.aau.dk>

**See Also**

[subSeq](#), [rle](#)

**Examples**

```
## Events:
yvar <- c(0, 0, 0, 1, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 1, 0, 1, 0, 0, 0,
         0, 0, 0, 0, 0, 1, 1, 0, 0, 0, 0, 0)

## Plot results:
tse<- timeSinceEvent(yvar)
plot(sign.tse~tvar, data=tse, type="b")
grid()
rug(tse$tvar[tse$yvar==1], col=4, lwd=4)
points(scale(tse$run), col=tse$run, lwd=2)
lines(abs.tse + .2 ~ tvar, data=tse, type="b", col=3)

## Find times for which time since an event is at most 1:
tse$tvar[tse$abs<=1]

yvar <- c(0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 1, 1, 0, 0, 0, 0,
         0, 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0)
)
tvar <- c(207, 208, 208, 208, 209, 209, 209, 209, 210, 210, 211, 211,
         211, 212, 213, 213, 214, 214, 215, 216, 216, 216, 216, 217, 217,
         217, 218, 218, 219, 219, 219, 219, 220, 220, 221, 221, 221, 221,
         222, 222, 222)

timeSinceEvent(yvar, tvar)
```

---

`which.maxn`*Where are the n largest or n smallest elements in a numeric vector ?*

---

**Description**

Determines the locations, i.e., indices of the n largest or n smallest elements of a numeric vector.

**Usage**

```
which.maxn(x, n = 1)
```

**Arguments**

<code>x</code>	numeric vector
<code>n</code>	integer $\geq 1$

**Value**

A vector of length at most n with the indices of the n largest / smaller elements. NAs are discarded and that can cause the vector to be smaller than n.

**Author(s)**

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**See Also**

[which.max](#), [which.min](#)

**Examples**

```
x <- c(1:4, 0:5, 11, NA, NA)
ii <- which.minn(x, 5)

x <- c(1, rep(NA,10), 2)
ii <- which.minn(x, 5)
```

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